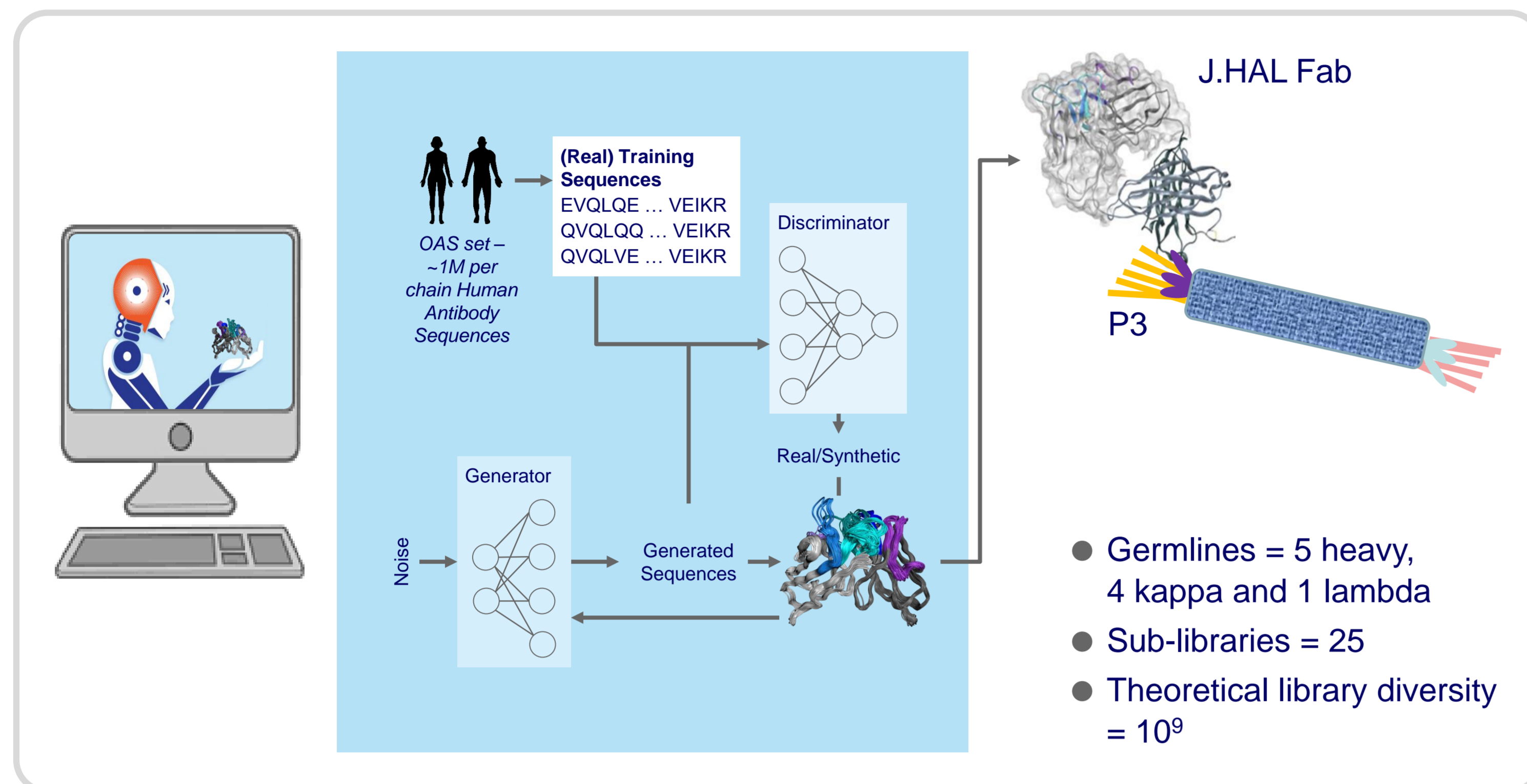


# AI-Derived Antibodies are novel, diverse and pharmacologically active against multiple SARS-CoV-2 strains

Cristina Moldovan Loomis<sup>1</sup>, Megan Sprague<sup>1</sup>, Andrew Asakawa<sup>1</sup>, Gregory Neveu<sup>2</sup>, Antoine Alam<sup>2</sup>, Randal Ketchum<sup>1</sup> and Rutilio Clark<sup>1</sup>  
<sup>1</sup>Just – Evotec Biologics Inc., US; <sup>2</sup>Evotec, France

We have developed an AI-generated antibody library platform utilizing a Generative Adversarial Network (GAN) that generates novel sequences which mimic natural human response, as well biasing toward diversity and developability features. The resulting Humanoid Antibody Library (J.HAL<sup>SM</sup>) was successfully screened to obtain a panel of novel, diverse and pharmacologically active human antibodies against SARS-CoV-2.

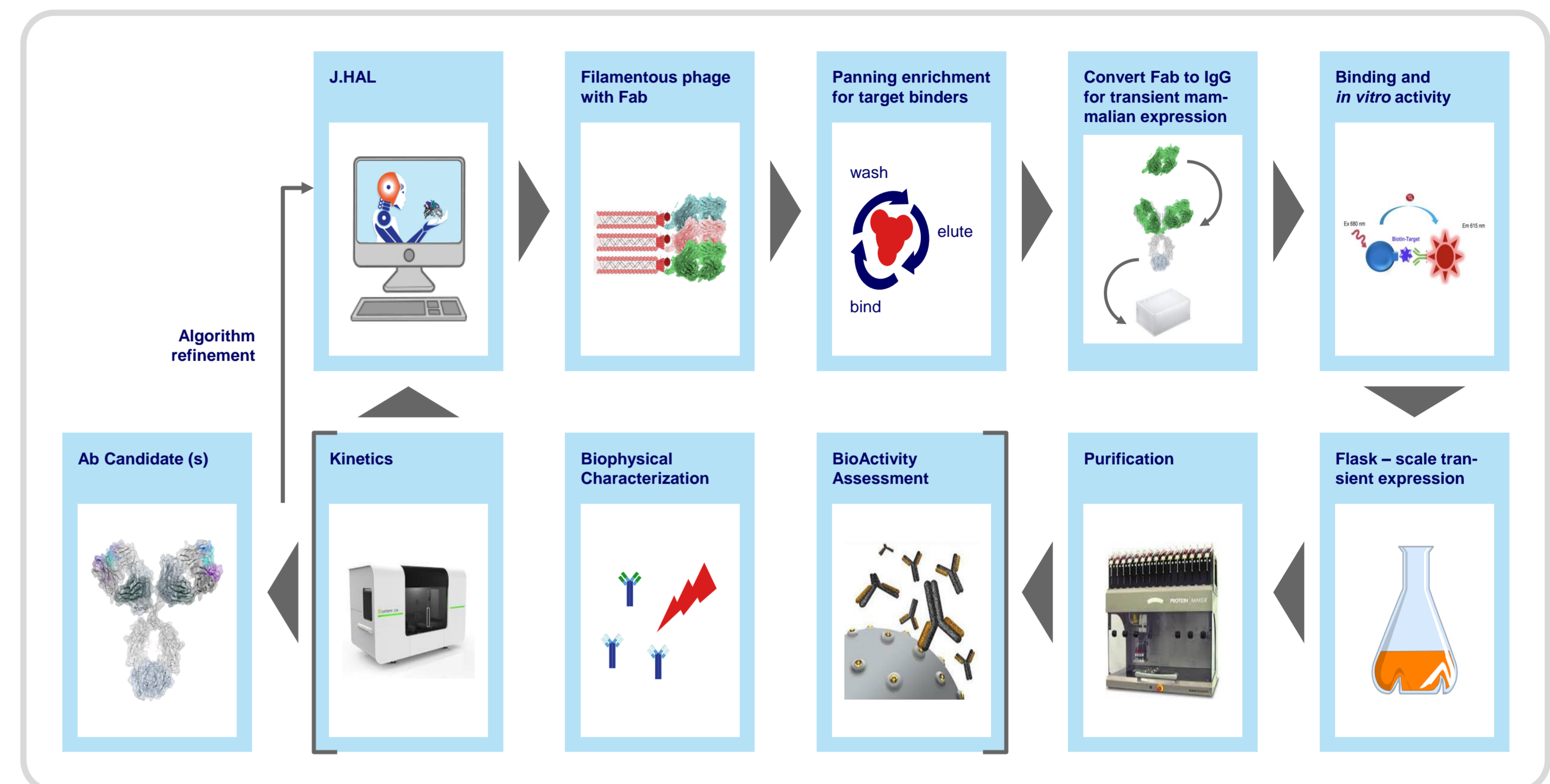
## J.HAL technology is a GAN application for antibody sequences



- Trained on **real** mature human antibody sequences and built as a phage library
- Large, human-derived antibody sequence training set extracted from OAS
- GAN training models are germline specific
- Ability to generate synthetic humanoid large, diverse, combinatorial germline pairings for library creation
- GAN-generated antibodies represent **B-cell response – including full SHM**

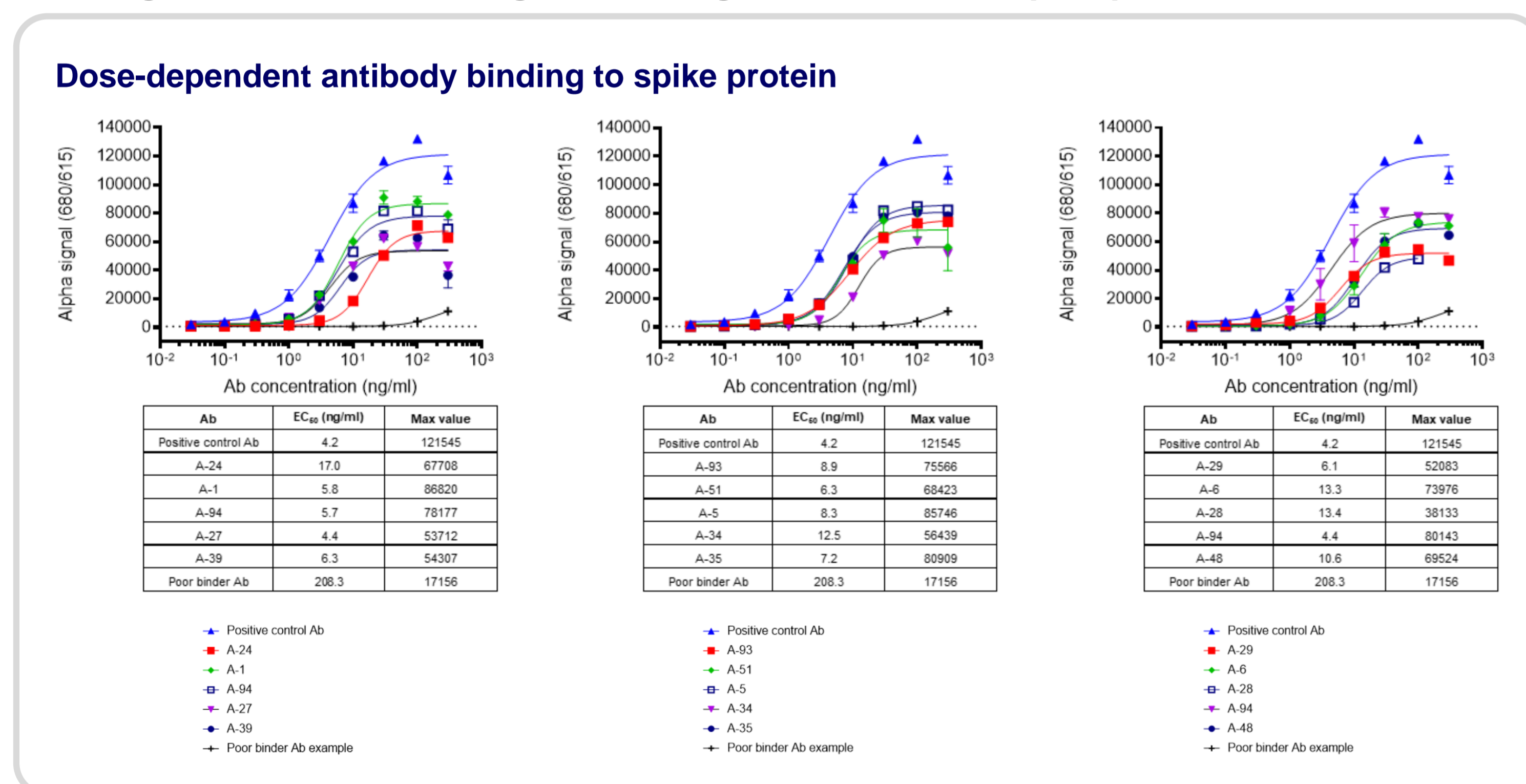
Preprint available at bioRxiv (<https://www.biorxiv.org/content/10.1101/2020.04.12.024844v2>)

## SARS-CoV-2 Antibody Discovery Workflow



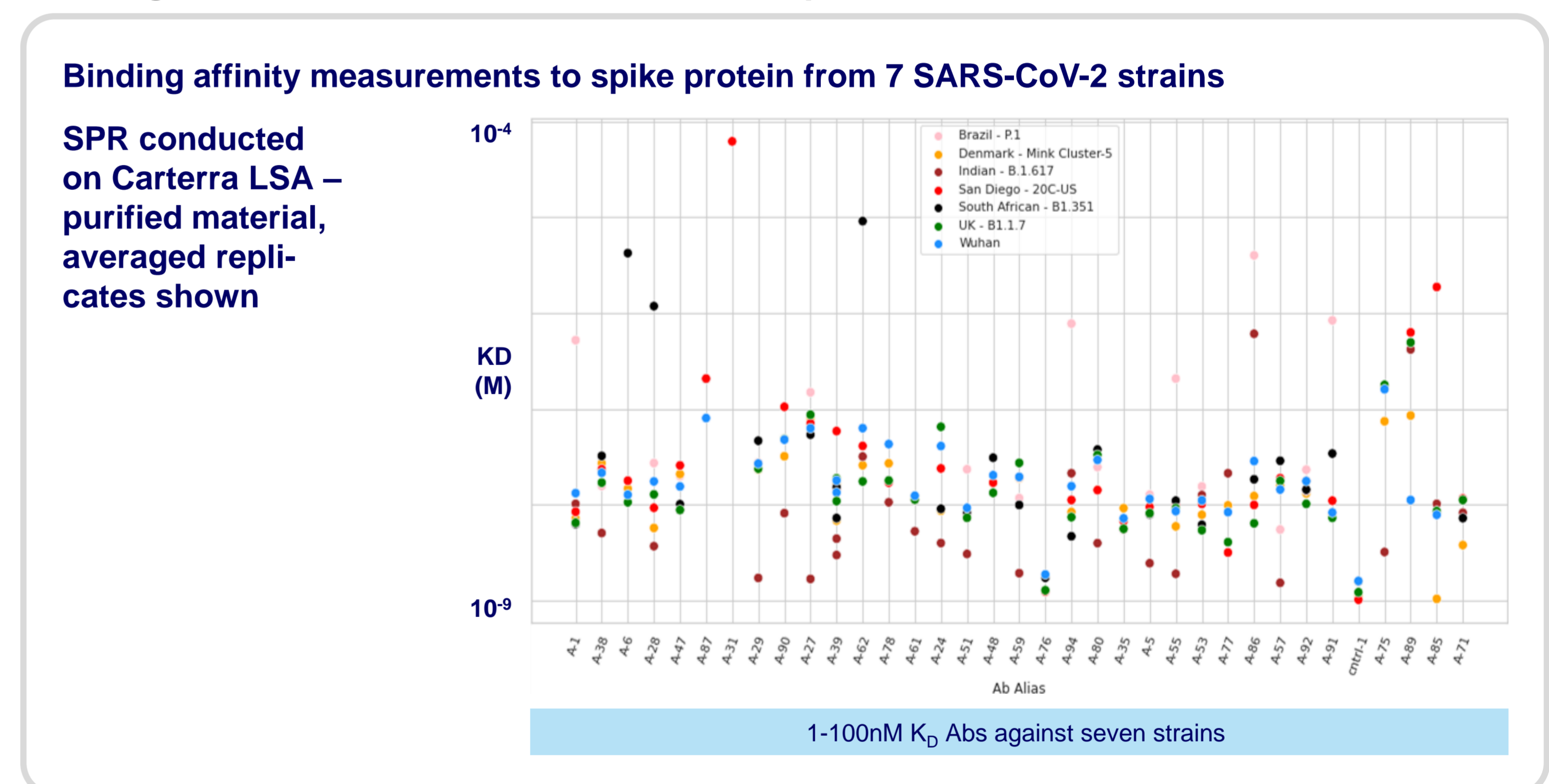
- J.HAL Fab on phage panned on Wuhan RBD or on UK S1 antigens with increasing stringency
- Enriched Fab converted to IgG and transiently expressed in 293F cells at 96 deep well scale
- Unpurified transfection supernatants used for screening binding and activity assays
- Top candidates expressed at flask scale, purified and tested for SARS-CoV-2 neutralization ability across multiple strains

## J.HAL IgG antibodies exhibit good binding to SARS-CoV-2 spike protein



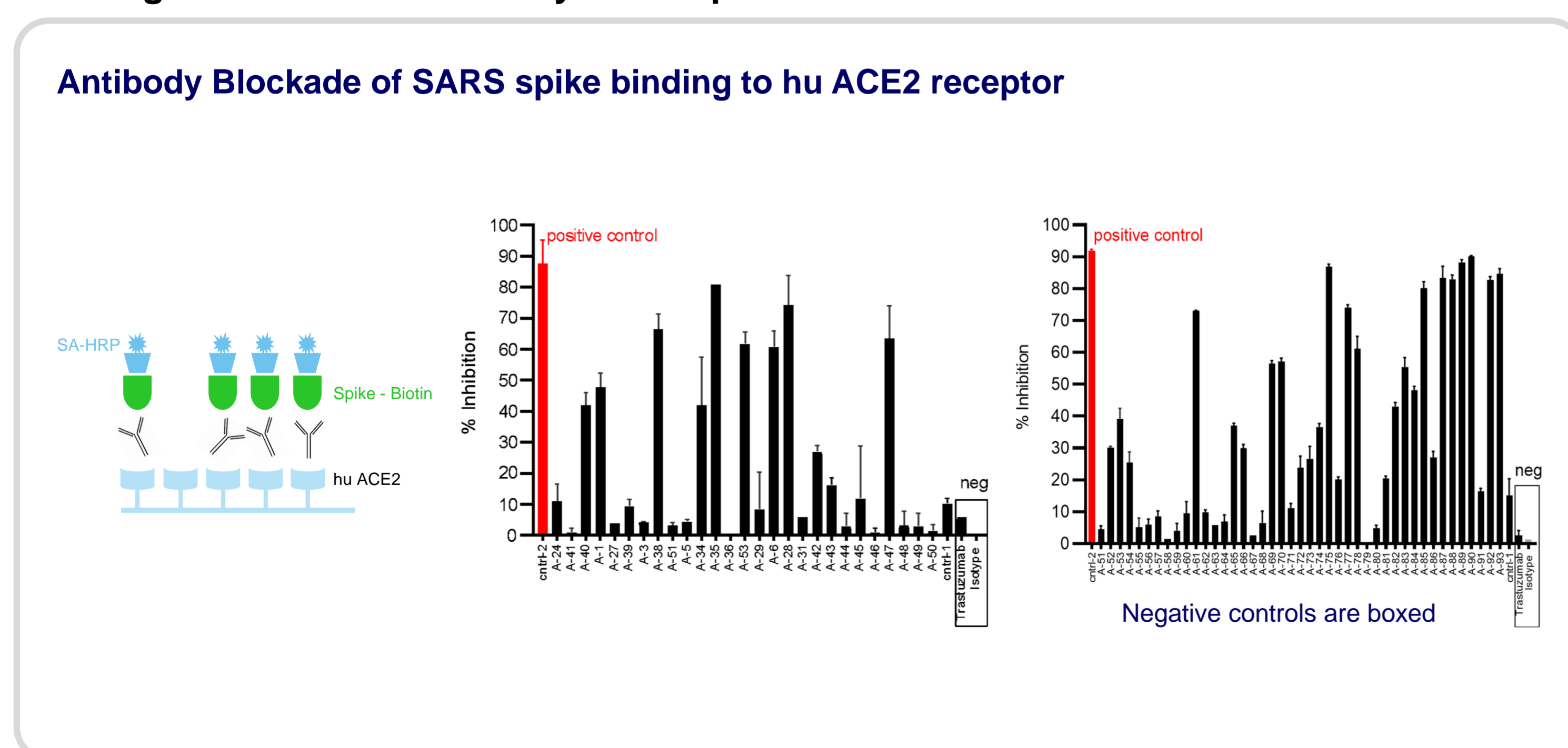
- Candidates that specifically bound SARS-CoV-2 spike protein and did not bind an irrelevant antigen were further characterized for dose-dependent binding using AlphaLISA technology
- A total of 73 unique antibody sequences specific for SARS-CoV-2 Spike protein were identified in the primary “yes/no” binding screen
- Binding assays were performed using unpurified transfection supernatants and later reproduced with purified material
- All antibody data are from **native library candidates without any affinity maturation**
- **Feasible to screen unpurified transfection supernatants to accelerate discovery timelines**

## J.HAL IgG antibodies cross-react across multiple SARS-CoV-2 strains



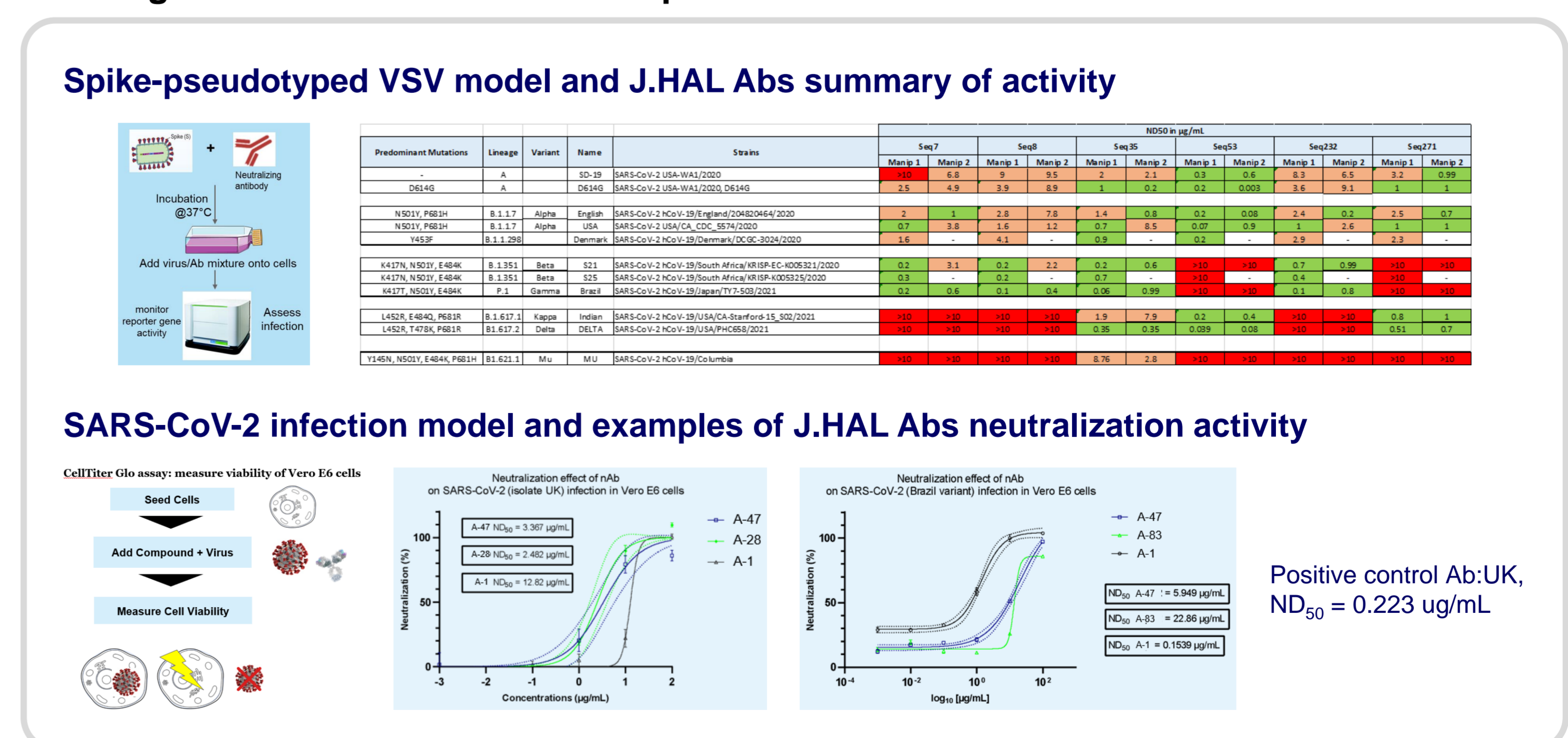
- Binding of selected candidates to the spike protein from seven viral strains was characterized by SPR conducted on Catterra LSA
- SPR binding profiles of purified candidates matched the binding profiles of unpurified transient transfection supernatants
- All antibody data are from **native library candidates without any affinity maturation**
- **Feasible to utilize SPR technology to assess pan cross-reactivity of unpurified transfection supernatants to accelerate discovery timelines**

## J.HAL IgG antibodies effectively block spike:huACE2R interaction



- Candidate antibody supernatants that specifically bound SARS-CoV-2 Spike protein were tested for their ability to block binding of SARS-CoV-2 spike protein to human ACE-2 receptor
- Multiple antibodies identified that effectively block spike: human ACE2 receptor interaction
- **Feasible to screen unpurified transfection supernatants for functional activity**

## J.HAL IgG antibodies neutralize multiple strains of SARS-CoV-2



- Neutralization ability of candidate antibodies was assessed using VSV pseudotyped-system harboring the spike envelope glycoprotein of SARS-CoV-2; multiple strains were evaluated
- Multiple candidates were demonstrated to have neutralizing activity against several strains of SARS-CoV-2
- All antibody data are from **native library candidates without any affinity maturation**

**Acknowledgements:** Danielle van Citters, Kathryn McLean, Lindsay Pautsch, Caroline Carbonelle, Fred Hutch Carterra LSA SPR facility